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Exploring causality with biliary atresia at different levels: two-sample Mendelian randomization study

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ABSTRACT

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Correspondence to Dr Jianghua Zhan; zhanjianghuatj@163.com **Background** In recent years, Mendelian randomization (MR) has been widely used to infer causality of related disease risk exposures. However, this strategy has not been applied to biliary atresia (BA).

Methods Genome-wide association studies (GWAS) data of 41 inflammatory cytokines, 731 immune cell traits, and 1400 metabolites were obtained from public databases as exposure factors. The outcome information was obtained from a GWAS meta-analysis of 499 children with BA and 1928 normal controls. Inverse variance weighting was the primary causality analysis. Cochran Q-test, MR-Egger intercept, MR pleiotropy residual sum and outlier, and 'leave-one-out' analyses were used for sensitivity analysis. Reverse MR, MR-Steiger, and Linkage Disequilibrium Score were used to exclude the effects of reverse causality, genetic association, and linkage disequilibrium. Results MR results showed that a total of seven traits had potential causal relationships with BA, including three inflammatory cytokines: eotaxin (odds ratio (OR)=1.45, 95% confidence interval (CI): 1.08 to 1.95, p_{FDR} =0.18), G-CSF (OR=4.21, 95% CI: 1.75 to 10.13, $p_{FDR} = 0.05$) and MCP-1/MCAF (OR=1.53, 95% CI: 1.12 to 2.10, p_{FDR}=0.14); three immune cell traits: CD8dim NKT/T cells ratio (0R=0.59, 95% CI: 0.45 to 0.77, p_{FDR}=0.06), CD8dim NKT counts (OR=0.58, 95% CI: 0.43 to 0.78, p_{FDR}=0.06), CD8dim NKT/lymphocyte ratio (OR=0.63, 95% CI: 0.49 to 0.81, p_{FDR} =0.06); one metabolite: X-12261 levels (OR=2.86, 95% CI: 1.73 to 4.74, p_{FDR}=0.06).

Conclusions In this study, eotaxin, G-CSF, MCP-1/MCAF, and X-12261 levels were shown to be risk factors for BA. However, CD8dim NKT/T cells ratio, CD8dim NKT counts, and CD8dim NKT/lymphocyte ratio were protective factors for BA. These findings provided a promising genetic basis for the etiology, diagnosis, and treatment of BA.

INTRODUCTION

Biliary atresia (BA) is one of the most serious hepatobiliary diseases in neonates, which is characterized by progressive inflammatory fibrosis and occlusion of intrahepatic and extrahepatic bile ducts.¹ The etiology of BA is still unclear, and the possible theoretical mechanisms include genetic variation, toxins, viral infection, autoimmune-mediated chronic inflammation or bile duct lesions,

WHAT IS ALREADY KNOWN ON THIS TOPIC

- ⇒ Biliary atresia (BA) exhibits inflammatory pathological features, and its occurrence may be closely associated with autoimmunity.
- ⇒ Previous research confirmed the possibility of metabonomics as a non-invasive biomarker for the early detection of BA.
- ⇒ Mendelian randomization (MR) analysis is a reliable causal assessment method, which has been widely used in various disease fields to find the possible causes of the disease. However, no study has reported the application of MR analysis in BA.

WHAT THIS STUDY ADDS

- $\Rightarrow\,$ The first MR analysis to explore causal relationships with BA from multiple levels.
- ⇒ Eotaxin, G-CSF, MCP-1/MCAF, and X-12261 were risk factors for BA. CD8dim NKT/T cell ratio, CD8dim NKT count, and CD8dim NKT/lymphocyte ratio were protective factors for BA.

HOW THIS STUDY MIGHT AFFECT RESEARCH, PRACTICE OR POLICY

⇒ This study conducted the MR analysis of BA from three different levels (inflammatory factors, immune cell traits, and metabolites), which would aid in understanding the pathogenesis of BA and provide a genetic basis for its diagnosis and treatment.

and abnormal bile duct development.² Early diagnosis of BA can effectively improve the prognosis of children, and MMP-7 is the most promising early diagnostic index of BA; due to the lack of unified criteria, its clinical application is limited.^{3 4} In addition, there is currently no effective drug for the treatment of BA. Therefore, there is an urgent need to find the etiology of BA, new early diagnostic markers, and therapeutic targets.

In recent years, Mendelian randomization (MR) has been widely used to infer causality of related disease risk exposures. This process is done using publicly available genome-wide association studies (GWAS) datasets. Because genotypes are determined at conception, MR

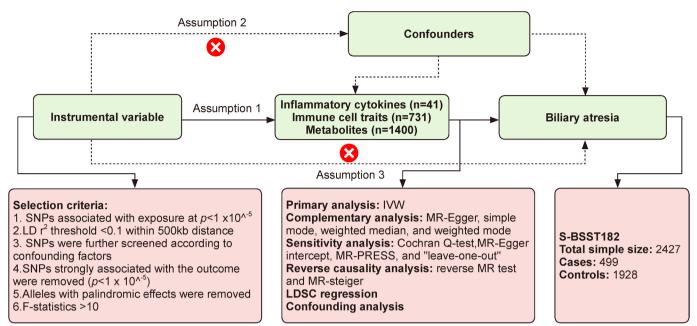


Figure 1 Flow chart of the Mendelian randomization (MR) study. IVW, inverse variance weighting; LD, linkage disequilibrium; LDSC, Linkage Disequilibrium Score; MR-PRESSO, MR pleiotropy residual sum and outlier; SNPs, single nucleotide polymorphisms.

analyses are generally not susceptible to confounding factors.

In conclusion, BA exhibits inflammatory pathological features, and its occurrence may be closely associated with autoimmunity. In addition, previous research confirmed the possibility of metabonomics as a noninvasive biomarker for the early detection of BA.⁵ Therefore, this study conducted the MR analysis of BA from three different levels (inflammatory factors, immune cell traits, and metabolites), which would aid in understanding the pathogenesis of BA and provide a genetic basis for its diagnosis and treatment.

METHODS

Research design

Based on the two-sample MR analysis, we assessed 41 inflammatory cytokines, 731 immune cell traits, and 1400 metabolites for their potential causal relationships with BA as the outcome. The instrumental variables (IVs) used in MR analysis must adhere to three assumptions: (1) IVs must be exposure related, (2) IVs must be independent of other confounders, (3) IVs are unrelated to the outcome and can influence the outcome only through exposure.⁶ Figure 1 provides an overview of this MR study, and the study methods align with the Strengthening the Reporting of Observational Studies in Epidemiology-MR checklist.⁷

GWAS data sources

We identified genetic predictors of 41 inflammatory cytokines from a comprehensive cytokine-related GWAS meta-analysis involving three independent cohorts. The study encompassed 8293 Finnish participants from the Cardiovascular Risk in Young Finns Study and the 'FINRISK' Studies (FINRISK1997 and FINRISK2002). 8

Each immune trait from the GWAS data can be accessed publicly via the GWAS Catalog (https://www.ebi.ac.uk/ gwas/, GCST0001391-GCST0002121).⁹ A total of 731 immune cell traits, comprising median fluorescence intensities reflecting surface antigen levels (n=389), relative cell (RC) counts (n=192), absolute cell (AC) counts (n=118), and morphological parameters (n=32), were included (online supplemental table 1). The GWAS for this immunologic profile used data from 3757 European individuals with no overlapping cohorts.

The GWAS dataset for 1400 metabolites was sourced from the study conducted by Chen et al.¹⁰ This represented the most comprehensive analysis of human metabolites, with complete summary statistics available through the GWAS Catalog (https://www.ebi.ac.uk/ gwas/, GCST90199621-GCST90201020) in the public domain. The extensive GWAS involved 1091 metabolites and 309 metabolite ratios in 8299 individuals from the Canadian Longitudinal Study on Aging cohort. The dataset comprises 1091 metabolites, including 850 known metabolites and 241 unknown metabolites (online supplemental table 2). These known metabolites, categorized into peptide, nucleotide, amino acid, carbohydrates, cofactors and vitamin, energy, lipid, and xenobiotics metabolism, are based on the Kyoto Encyclopedia of Genes and Genomes database.

The GWAS dataset for BA was obtained from the study conducted by Chen *et al.*¹¹ The first GWAS was conducted in a European-American cohort comprising 343 patients with isolated BA and 1716 controls. The second GWAS involved a European-American cohort with 156 patients

presenting with BA and other extrahepatic abnormalities, alongside 212 controls. GWAS summary data can be accessed at https://www.ebi.ac.uk/biostudies/ (S-BSST182).

Selection of IVs

The IVs selected for this MR analysis are based on three fundamental assumptions. Initially, single nucleotide polymorphisms (SNPs) with association thresholds at $p < 1 \times 10^{-5}$ were extracted for each exposure. Subsequently, the clumping program within PLINK software (V.1.90) was employed to prune these SNPs, using a linkage disequilibrium (LD) r^2 threshold of <0.1 within a 500 kb distance. The LD r² was calculated based on the 1000 Genomes Project as a reference panel.¹² Third, to validate the second hypothesis, we employed the 'phenoscanner' function to control for confounding factors. Fourth, to ensure the validity of the third hypothesis, we excluded SNPs strongly associated with the outcome variable $(p < 1 \times 10^{-5})$. Additionally, alleles with palindromic effects were removed, and the explained variance (\mathbf{R}^2) and F-statistics parameters were used to assess the strength of IVs. A threshold of F-statistics >10 is generally recommended for MR analysis. Finally, we conducted MR analysis on metabolites comprising more than two SNPs, calculating \mathbf{R}^2 and F-statistics as follows:

$$R^{2} = \frac{2 \times \beta^{2} \times EAF \times (1 - EAF)}{\left[2 \times \beta^{2} \times EAF \times (1 - EAF) + 2 \times (se(\beta))^{2} \times N \times EAF \times (1 - EAF)\right]}$$
$$F = \frac{N - k - 1}{k} \times \frac{R^{2}}{1 - R^{2}}$$

Where β represents the effect size of the genetic variant of interest; EAF denotes the effect allele frequency of the genetic variant of interest; se (β) indicates the SE of the effect size of the genetic variant of interest; N stands for the exposure sample size; and k is the number of SNPs.

MR analysis

This MR analysis primarily estimated the causal association using standard inverse variance weighting (IVW) methods.¹³ When the IVs satisfy all three assumptions of the valid IVs, the IVW approach can provide a consistent assessment of the causality of the exposure. However, if some IVs deviate from these assumptions, the analysis may yield inaccurate results. Hence, the following analyses were conducted: (1) heterogeneity among selected IVs was assessed using the Cochran Q-test with the corresponding p value. If the null hypothesis was rejected, a random-effect IVW was applied instead of a fixed-effect IVW.¹⁴ (2) Horizontal pleiotropy was evaluated through the intercept using the MR-Egger method to ensure that genetic variants were independently associated with both exposure and outcome.¹⁵ (3) Additional analyses using MR methods with different modeling assumptions and strengths (MR-Egger, simple mode, weighted median^{1b}, and weighted mode¹⁷) were performed to increase the stability and robustness of the results. (4) The MR pleiotropy residual sum and outlier (MR-PRESSO) method

was used to identify and exclude horizontal pleiotropic outliers that might significantly influence the estimated results.¹⁸ (5) A 'leave-one-out' sensitivity analysis was employed to assess whether the results were influenced by any single SNP.¹⁵ Additionally, we conducted MR-Steiger and reverse MR tests to ascertain whether our results supported our hypothesis.

Evaluation of genetic correlation and directionality

Although SNPs associated with BA were excluded in the selection of IVs, unrelated SNPs might have mediated the inheritance of BA. Therefore, to ensure that causal effects were not confounded by exposure consistency with the outcome, the SNP-based Linkage Disequilibrium Score (LDSC) was used to estimate coinheritance by X^2 statistics for both traits.

Confounding analysis

Although we evaluated horizontal pleiotropy in the MR results through various sensitivity analyses to detect SNPs violating the MR assumption, residual confounding SNPs might still exist. The 'phenoscanner' function was employed to assess whether each SNP was associated with known risk factors for BA, such as viral infection and maternal exposure to certain chemicals.¹⁹ If any SNP was found to be associated with the mentioned confounders ($p<1\times10^{-5}$), the MR analysis was re-executed after excluding these SNPs to validate the reliability of the results.

Statistical analysis

Statistical analyses were conducted using R V.4.1.2 software (http://www.Rproject.org). MR analysis was carried out using the 'TwoSampleMR' package (V.0.5.7).²⁰ IVW, weighted median-based, and model-based approaches were performed with the use of the 'MendelianRandomization' package (V.0.7.0).²¹ The false discovery rate (FDR)-adjusted p value, below 0.2, suggests statistical significance.²²

RESULTS

Causal assessment of 41 inflammatory cytokines for BA

To investigate the causal relationship between inflammatory cytokines and BA, a two-sample MR analysis was conducted. Using IVW as the primary method, three suggestive associations were detected following FDR adjustment (p_{FDR} <0.2); we identified three inflammatory cytokines as risk factors for the development of BA (figure 2A): eotaxin (odds ratio (OR)=1.45, 95% confidence interval (CI): 1.08 to 1.95, p_{FDR} =0.18, p=0.014), G-CSF (OR=4.21, 95% CI: 1.75 to 10.13, p_{FDR} =0.05, p=0.001) and MCP-1/MCAF (OR=1.53, 95% CI: 1.12 to 2.10, p_{FDR} =0.14, p=0.007).

Causal assessment of 731 immune cell traits for BA

To investigate the causal relationship between immune cell traits and BA, a two-sample MR analysis was employed.

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Α						
Exposure	Method	nSNP		OR (95% CI)	Ρ	P_FDR
Eotaxin	MR Egger	22	k <mark></mark>	2.10 (0.91 - 4.87)	0.097	
	weighted median	22		1.68 (1.09 - 2.57)	0.017	
	Inverse variance weighted (fixed effects)	22	⊷ 1	1.45 (1.08 - 1.95)	0.014	0.18
	Weighted mode	22		1.64 (0.98 - 2.72)	0.072	
	Simple mode	22	·····	1.48 (0.74 - 2.98)	0.282	
G-CSF	MR Egger	5		▶ 6.87 (0.23 - 207.09)	0.348	
	Weighted median	5		▶ 4.60 (1.57 - 13.49)	0.005	
	Inverse variance weighted (fixed effects)	5	· · · · ·	▶ 4.21 (1.75 - 10.13)	0.001	0.05
	Weighted mode	5	· · · · · · · · · · · · · · · · · · ·	> 5.72 (1.36 - 24.15)	0.076	
	Simple mode	5		> 5.75 (1.30 - 25.29)	0.082	
MCP-1/MCAF	MR Egger	23		2.71 (1.21 - 6.09)	0.025	
	Weighted median	23)	1.67 (1.03 - 2.69)	0.036	
	Inverse variance weighted (fixed effects)	23		1.53 (1.12 - 2.10)	0.007	0.14
	Weighted mode	23		1.78 (1.00 - 3.15)	0.063	
	Simple mode	23	H H	0.94 (0.41 - 2.14)	0.883	
			0 1 3	8		
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Protective factors for BA Risk factors for BA

В						
Exposure	Method	nSNP		OR (95% CI)	Ρ	P_FDR
CD8dim NKT/T cell ratio	MR Egger	17	H	0.40 (0.20 - 0.81)	0.02	
	Weighted median	17	—	0.54 (0.36 - 0.81)	0.00	
	Inverse variance weighted (fixed effects) 17	⊢ −−−1	0.59 (0.45 - 0.77)	0.00	0.06
	Weighted mode	17	→	0.47 (0.28 - 0.79)	0.01	
	Simple mode	17	► • • • • • •	0.44 (0.22 - 0.89)	0.04	
CD8dim NKT counts	MR Egger	19	H	0.26 (0.07 - 0.97)	0.06	
	Weighted median	19	⊢ •−−−1	0.55 (0.36 - 0.84)	0.01	
	Inverse variance weighted (fixed effects) 19	⊢ →−→	0.58 (0.43 - 0.78)	0.00	0.06
	Weighted mode	19		0.44 (0.21 - 0.90)	0.04	
	Simple mode	19	—	0.47 (0.21 - 1.05)	0.08	
CD8dim NKT/lymphocyte ratio	MR Egger	19	· · · · · · · · · · · · · · · · · · ·	0.62 (0.31 - 1.23)	0.19	
	Weighted median	19	⊢ •−−1	0.56 (0.37 - 0.83)	0.00	
	Inverse variance weighted (fixed effects) 19	I	0.63 (0.49 - 0.81)	0.00	0.06
	Weighted mode	19		0.49 (0.27 - 0.87)	0.03	
	Simple mode	19		0.47 (0.21 - 1.04)	0.08	
			0 0.5 1	1.5		
		\leftarrow				



Exposure	Method	nSNP		OR (95% CI)	Ρ	P_FDR
X-12261 levels	MR Egger	9	⊢ ¦ <mark>- </mark>	1.45 (0.12 - 17.20)	0.78	
	Weighted median	9	·•	2.37 (1.15 - 4.89)	0.02	
	Inverse variance weighted (fixed effects)	9		2.86 (1.73 - 4.74)	0.00	0.06
	Weighted mode	9		1.93 (0.61 - 6.11)	0.29	
	Simple mode	9	H	1.85 (0.55 - 6.28)	0.35	
			0 1 4	7		
	←			•		

Protective factors for BA Risk factors for BA

Figure 2 Forest plots of positive results from Mendelian randomization (MR) analysis. (A) Inflammatory cytokines, (B) immune cell traits and (C) metabolites. BA, biliary atresia; FDR, false discovery rate; SNP, single nucleotide polymorphism.

Using IVW as the primary analysis method, a total of 34 suggestive associations were identified, following FDR adjustment (p_{FDR} <0.2); we identified three immune cell traits as risk factors for the development of BA, two of

which were derived from RC counts and one from AC counts (figure 2B): CD8dim NKT/T cells ratio (OR=0.59, 95% CI: 0.45 to 0.77, p_{FDR} =0.06, p=0.000), CD8dim NKT counts (OR=0.58, 95% CI: 0.43 to 0.78, p_{FDR} =0.06,

Causal assessment of 1400 metabolites for BA To investigate the causal relationship between metabolites and BA, a two-sample MR analysis was conducted. Employing IVW as the primary analysis method, a total of 78 suggestive associations were identified following FDR adjustment ($p_{FDR} < 0.2$); we identified only one metabolite as a risk factor for BA development, and it came from the category of unknown metabolites (figure 2C): X-12261 levels (OR=2.86, 95% CI: 1.73 to 4.74, p_{FDR} =0.06, p=0.000). The detailed data of IVs were shown in online supplemental table 3. Generally, when statistical significance was observed in two other MR tests (p<0.05), the causal association is deemed robust. Notably, the association of MCP-1/MCAF, CD8dim NKT counts, CD8dim NKT/T cells ratio, CD8dim NKT/lymphocyte ratio, and X-12261 levels was deemed robust (online supplemental table 4). Although eotaxin and G-CSF were significant only in the IVW and weighted median methods (p < 0.05), they still

Heterogeneity test

mental table 4).

Heterogeneity was tested using the Cochran Q-test. The p values of seven traits (eotaxin, G-CSF, MCP-1/MCAF, CD8dim NKT count, CD8dim NKT/T cell ratio, CD8dim NKT/lymphocyte ratio, and X-12261 level) were greater than 0.05, indicating that no significant heterogeneity was found (online supplemental table 4).

signified potential causal associations (online supple-

p=0.000), CD8dim NKT/lymphocyte ratio (OR=0.63,

95% CI: 0.49 to 0.81, $p_{\mu\nu\rho}=0.06$, p=0.000).

Sensitivity analysis

Sensitivity analyses were performed to identify horizontal pleiotropy and potential outliers. All associations were scrutinized for potential horizontal pleiotropy using the MR-Egger intercept and MR-PRESSO global test. All p values in the results were greater than 0.05, indicating that there was no significant horizontal pleiotropy (online supplemental table 4). Additionally, we visualized scatter plots (figure 3), funnel plots (online supplemental figure 1), and 'leave-one-out' plot (online supplemental figure 2) from which no potential outliers and the possibility of horizontal pleiotropy were found.

Reverse causality analysis

To test hypothesis 3 ('IVs are unrelated to the outcome and can only influence the outcome through exposure'), MR-Steiger and reverse MR tests were conducted. Results indicated that in reverse causality analysis with BA as the exposure factor, the p values of the MR-Steiger for eotaxin, G-CSF, MCP-1/MCAF, and X-12261 levels were all below 0.05. And their p values of the reverse MR test were all above 0.05, confirming the study's correct directionality (online supplemental tables 4 and 5). However, due to the lack of data for immune cell traits in the reverse MR test, their association with BA was verified solely through the MR-Steiger method. CD8dim NKT counts, CD8dim NKT/T cells ratio, and CD8dim NKT/ lymphocyte ratio all had the p values of the MR-Steiger below 0.05, validating the study's directionality (online supplemental table 4).

Linkage Disequilibrium Score

The results of LDSC showed that eotaxin (R =0.068, SE=0.110, p=0.537), G-CSF (R =0.572, SE=0.313, p=0.068), MCP-1/MCAF (R =-0.077, SE=0.081, p=0.342), and X-12261 levels (R =-0.060, SE=0.161, p=0.711), which indicated that the MR estimates were not disturbed by the shared genetic component (table 1). We estimated the SNP–SNP heritability (proportion of variance attributed to genome-wide SNPs) for the four associations using LDSC. The SNP heritability of metabolites varied from 0.015 (G-CSF) to 0.167 (MCP-1/MCAF) (online supplemental table 6). The immune cell traits dataset was not analyzed as it lacked the necessary data for LDSC completion.

Confounding analysis

Although sensitivity analysis in this study was conducted to assess SNP estimates, to meet assumption 2 ('IVs must be independent of other confounders'), we examined whether all SNPs associated with the seven traits were independent of BA risk factors (viral infection and maternal exposure to certain chemicals). Results indicated that none of the seven traits were associated with any confounding factors (online supplemental table 7).

DISCUSSION

Based on extensive publicly available genetic data, we investigated the causal relationship between various factors, including 41 inflammatory cytokines, 731 immune cell traits, and 1400 metabolites, and the disease outcome of BA. To our knowledge, this is the first MR analysis to explore causal relationships with BA at multiple levels. Our study identified three inflammatory cytokines (eotaxin, G-CSF, MCP-1/MCAF), three immune cell traits (CD8dim NKT counts, CD8dim NKT/T cells ratio, CD8dim NKT/lymphocyte ratio), and one metabolite (X-12261 levels) as having a significant causal relationship with BA.

The expression of each of the three inflammatory factors was a risk factor for BA, respectively, consistent with previous studies. Udomsinprasert *et al*²³ found that eotaxin, G-CSF, and MCP-1 levels were higher in the blood of children with BA. Increased eotaxin levels were notably linked to adverse outcomes such as jaundice, fibrosis, and portal hypertension. Moreover, the study identified MCP-1 as a potentially sensitive and specific biomarker for BA, with significantly upregulated relative mRNA expression observed in BA livers.

MCP-1 may play an important role in the development of BA with progressive liver fibrosis. Glucocorticoid treatment can regulate the expression of MCP-1 in the liver of cholestatic rats and reduce the infiltration

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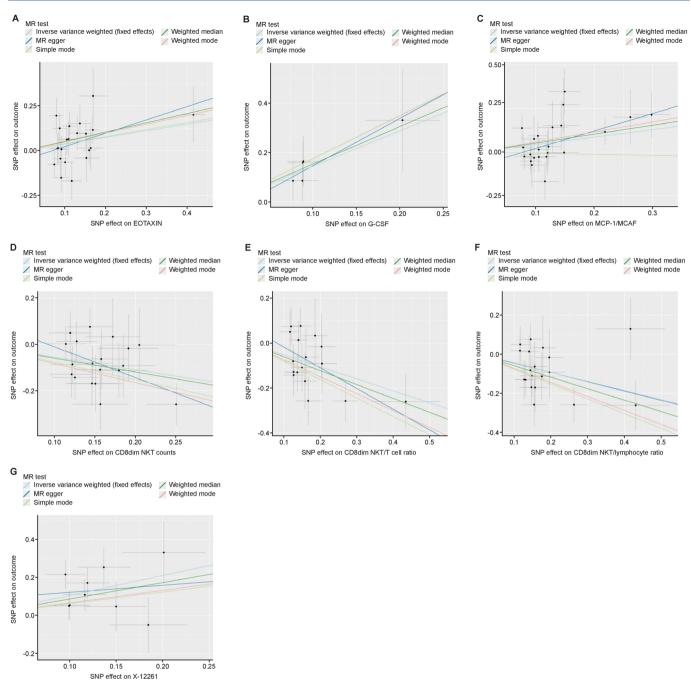


Figure 3 Scatter plots of positive results for Mendelian randomization (MR) analysis. (A) eotaxin, (B) G-CSF, (C) MCP-1/ MCAF, (D) CD8dim NKT counts, (E) CD8dim NKT/T cells ratio, (F) CD8dim NKT/lymphocyte ratio and (G) X-12261. SNP, single nucleotide polymorphism.

Table 1 Results of LDSC						
Trait1	Trait2	R _a	SE	P value		
Eotaxin	BA	0.068	0.110	0.537		
G-CSF	BA	0.572	0.313	0.068		
MCP-1/MCAF	BA	-0.077	0.081	0.342		
X-12261 levels	BA	0.060	0.161	0.711		
RA biliany atropia: LDSC Linkago Disoguilibrium Score						

BA, biliary atresia; LDSC, Linkage Disequilibrium Score.

of inflammatory cells.^{24 25} Ramm *et al*²⁶ found that bile acid-induced upregulation of hepatocyte-derived MCP-1 expression led to the recruitment of hepatic stellate cells and was a critical early event in the development of liver fibrosis. Jafri *et al*²⁷ found that chemokines expressed by rhesus rotavirus-infected cholangiocytes, such as MCP-1, might trigger host inflammatory processes leading to bile duct obstruction.

In a phase 1 study exploring Kasai-G-CSF sequential therapy for BA, G-CSF safely mobilized hematopoietic stem cells in Kasai children, potentially enhancing

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short-term biliary drainage and mitigating cholangitis.²⁸ Nonetheless, the efficacy outcomes of sequential adjuvant Kasai and G-CSF therapy in phase 2 have yet to be determined. Intriguingly, another study observed high expression of G-CSF in the liver tissues of children with BA.²³ While our findings suggest that G-CSF is a risk factor for BA, discrepancies between MR analysis and validation results might occur, influenced by factors such as insufficient sample size and small effect size. Hence, for G-CSF, we will further validate its relationship with BA in subsequent studies.

Our study revealed a decrease in the risk of BA with increasing CD8dim NKT counts, CD8dim NKT/T cells ratio, and CD8dim NKT/lymphocyte ratio. Notably, the association between CD8dim NKT counts and BA had not been previously reported. Previous research on the connection between BA and CD8+ T cells showed that a higher degree of CD8+ T cell infiltration in the bile duct of children with BA was correlated with better liver function.²⁹ In addition, Kotb *et al*⁸⁰ observed a greater mean CD4+/CD8+ ratio in children who died within eighteen months after the Kasai procedure than in other children with BA.

Our study has several strengths. First, the two-sample MR study design helps reduce bias present in observational association studies, stemming from residual confounding and reverse causality. Second, we systematically investigated the causal relationship between multiple levels of data (including inflammatory cytokines, immune cell traits, and metabolites) and BA. Third, we employed seven MR methods to assess the robustness of causal associations and effect directions, including IVW, MR-Egger, weighted median, weighted mode, simple mode, MR-PRESSO, and MR-Steiger.

However, there are limitations to consider. First, we did not further subdivide BA considering the classification of the original data. Second, the power of IVs relies heavily on the sample size of GWAS, necessitating more data to enhance accuracy. Third, while MR analysis is reliable for assessing causality, it cannot replace randomized controlled trials (RCTs). Therefore, inferred causal relationships may not align with those observed in RCTs and require further validation in future studies. Fourth, our study relied on the Euro-American cohort for GWAS data, limiting the generalizability of our findings to other ethnic groups.

In conclusion, our study used publicly available GWAS data and MR analysis to identify three inflammatory cytokines (eotaxin, G-CSF, MCP-1/MCAF) and one metabolite X-12261 as risk factors for BA with a significant causal relationship. Three immune cell characteristics (CD8dim NKT counts, CD8dim NKT/T cells ratio, CD8dim NKT/ lymphocyte ratio) also had a significant causal relationship with BA, and they were protective factors for BA. These findings contributed to a new genetic understanding of BA's etiology, diagnosis, and potential treatment strategies. In our subsequent work, we will evaluate the accuracy of three inflammatory cytokines in the early diagnosis of BA using clinical samples. Furthermore, we will use in vivo and in vitro experiments along with multiomics techniques to further investigate the involvement of these three inflammatory cytokines and CD8dim NKT cells in the pathogenesis of BA, exploring their specific roles in disease progression.

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Contributors Conception and design—SL. Administrative support—JZ. The guarantor—JZ. Collection and assembly of data—SL, XW, and TL. Data analysis and interpretation—SL, JM, and ZW. Manuscript writing—all authors. Final approval of the manuscript—all authors. The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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Data availability statement Data are available in a public, open access repository. Data sources are included in the Methods section of the article. Further information is available from the corresponding author.

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REFERENCES

- 1 Nio M. Japanese biliary atresia registry. *Pediatr Surg Int* 2017;33:1319–25.
- 2 Quelhas P, Cerski C, Dos Santos JL. Update on etiology and pathogenesis of biliary atresia. *Curr Pediatr Rev* 2022;19:48–67.
- 3 Karbasian F, Mashhadiagha A, Anbardar MH, *et al.* Questioning diagnostic value of serum matrix metalloproteinase 7 for biliary atresia. *J Clin Exp Hepatol* 2023;13:265–72.
- 4 Lertudomphonwanit C, Mourya R, Fei L, et al. Large-scale proteomics identifies MMP-7 as a sentinel of epithelial injury and of biliary atresia. Sci Transl Med 2017;9:eaan8462.
- 5 Zhou K, Xie G, Wang J, et al. Metabonomics reveals metabolite changes in biliary atresia infants. J Proteome Res 2015;14:2569–74.
- 6 Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data.
- Genet Epidemiol 2013;37:658–65.
 7 Skrivankova VW, Richmond RC, Woolf BAR, et al. Strengthening the reporting of observational studies in epidemiology using mendelian randomization: the STROBE-MR statement. JAMA 2021;326:1614–21.
- 8 Ahola-Olli AV, Würtz P, Havulinna AS, et al. Genome-wide association study identifies 27 loci influencing concentrations of circulating cytokines and growth factors. Am J Hum Genet 2017;100:40–50.

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- 9 Orrù V, Steri M, Sidore C, et al. Complex genetic signatures in immune cells underlie autoimmunity and inform therapy. Nat Genet 2020;52:1036–45.
- 10 Chen Y, Lu T, Pettersson-Kymmer U, *et al.* Genomic atlas of the plasma metabolome prioritizes metabolites implicated in human diseases. *Nat Genet* 2023;55:44–53.
- 11 Chen Y, Gilbert MA, Grochowski CM, et al. A genome-wide association study identifies a susceptibility locus for biliary atresia on 2p16.1 within the gene EFEMP1. *PLoS Genet* 2018;14:e1007532.
- 12 Auton A, Brooks LD, Durbin RM, *et al.* A global reference for human genetic variation. *Nature* 2015;526:68–74.
- 13 Burgess S, Small DS, Thompson SG. A review of instrumental variable estimators for mendelian randomization. *Stat Methods Med Res* 2017;26:2333–55.
- 14 Cohen JF, Chalumeau M, Cohen R, et al. Cochran's Q test was useful to assess heterogeneity in likelihood ratios in studies of diagnostic accuracy. J Clin Epidemiol 2015;68:299–306.
- 15 Burgess S, Thompson SG. Interpreting findings from mendelian randomization using the MR-Egger method. *Eur J Epidemiol* 2017;32:377–89.
- 16 Hartwig FP, Davey Smith G, Bowden J. Robust inference in summary data mendelian randomization via the zero modal pleiotropy assumption. Int J Epidemiol 2017;46:1985–98.
- 17 Bowden J, Davey Smith G, Haycock PC, et al. Consistent estimation in mendelian randomization with some invalid instruments using a weighted median estimator. Genet Epidemiol 2016;40:304–14.
- 18 Verbanck M, Chen CY, Neale B, et al. Detection of widespread horizontal pleiotropy in causal relationships inferred from mendelian randomization between complex traits and diseases. *Nat Genet* 2018;50:693–8.
- 19 Lendahl U, Lui VCH, Chung PHY, *et al.* Biliary atresia emerging diagnostic and therapy opportunities. *EBioMedicine* 2021;74:103689.

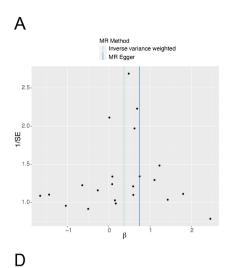
- 20 Hemani G, Zheng J, Elsworth B, et al. The MR-base platform supports systematic causal inference across the human phenome. *Elife* 2018;7:e34408.
- 21 Yavorska OO, Burgess S. Mendelianrandomization: an R package for performing mendelian randomization analyses using summarized data. *Int J Epidemiol* 2017;46:1734–9.
- 22 Wang C, Zhu D, Zhang D, et al. Causal role of immune cells in schizophrenia: mendelian randomization (MR) study. *BMC Psychiatry* 2023;23:590.
- 23 Udomsinprasert W, Ungsudechachai T, Vejchapipat P, et al. Systemic cytokine profiles in biliary atresia. *PLoS One* 2022;17:e0267363.
- 24 Hsieh CS, Huang CC, Huang LT, *et al*. Glucocorticoid treatment down-regulates chemokine expression of bacterial cholangitis in cholestatic rats. *J Pediatr Surg* 2004;39:10–5.
- 25 Kobayashi H, Tamatani T, Tamura T, et al. The role of monocyte chemoattractant protein-1 in biliary atresia. J Pediatr Surg 2006;41:1967–72.
- 26 Ramm GA, Shepherd RW, Hoskins AC, et al. Fibrogenesis in pediatric cholestatic liver disease: role of taurocholate and hepatocyte-derived monocyte chemotaxis protein-1 in hepatic stellate cell recruitment. *Hepatology* 2009;49:533–44.
- 27 Jafri M, Donnelly B, Bondoc A, et al. Cholangiocyte secretion of chemokines in experimental biliary atresia. J Pediatr Surg 2009;44:500–7.
- 28 Holterman A, Nguyen HPA, Nadler E, et al. Granulocyte-colony stimulating factor GCSF mobilizes hematopoietic stem cells in kasai patients with biliary atresia in a phase 1 study and improves short term outcome. J Pediatr Surg 2021;56:1179–85.
- 29 Ahmed AF, Ohtani H, Nio M, et al. Cd8+ T cells infiltrating into bile ducts in biliary atresia do not appear to function as cytotoxic T cells: a clinicopathological analysis. J Pathol 2001;193:383–9.
- 30 Kotb MA, El Henawy A, Talaat S, et al. Immune-mediated liver injury: prognostic value of CD4+, CD8+, and CD68+ in infants with extrahepatic biliary atresia. J Pediatr Surg 2005;40:1252–7.

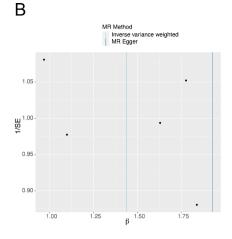
Supplementary Figure 1 Funnel plots of positive results for Mendelian randomization analysis. A Eotaxin. B G-CSF. C MCP-1/MCAF. D CD8dim NKT counts. E CD8dim NKT/T cells ratio. F CD8dim NKT/lymphocyte ratio.

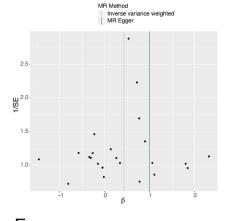
Supplementary Figure 2 "leave-one-out" analyses of positive results for Mendelian randomization analysis. A

Eotaxin. B G-CSF. C MCP-1/MCAF. D CD8dim NKT counts. E CD8dim NKT/T cells ratio. F CD8dim

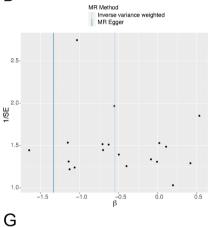
NKT/lymphocyte ratio.



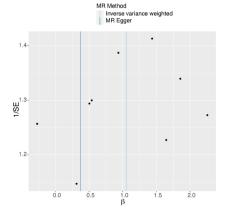


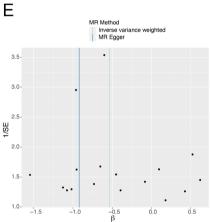


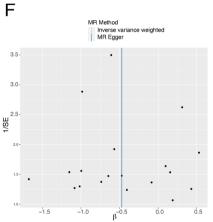
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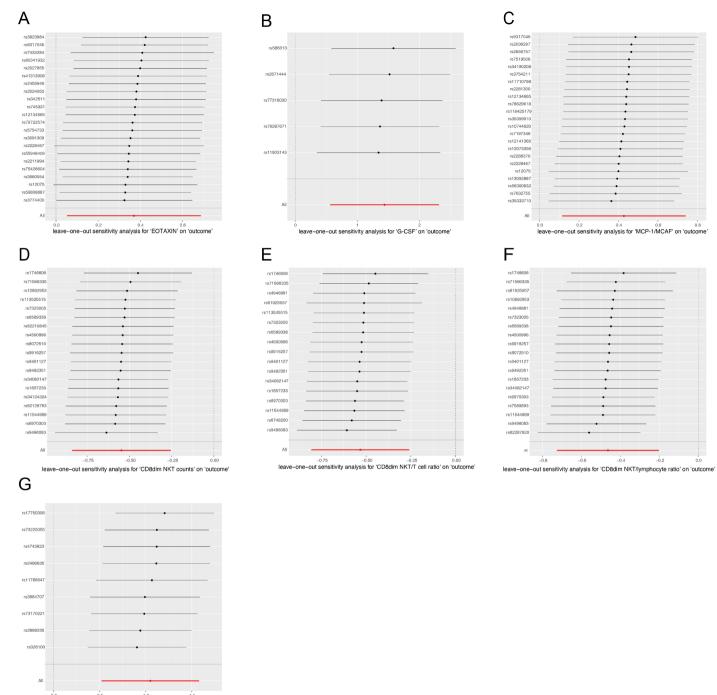












leave-one-out sensitivity analysis for 'X-12261' on 'outcome'